Morphological and Biochemical Characterization of Wild Mushrooms from Sub Mountainous Region of Punjab State

Amanpreet Kaur¹ and Dr. H.S. Sodhi²

¹Ph. D. Scholar, Department of Microbiology Punjab Agricultural University, Ludhiana
²Sr. Mycologist, Department of Microbiology Punjab Agricultural University, Ludhiana E-mail: ¹preetamanpau@gmail.com, ²drhssodhig@gmail.com

Abstract—The diversity of wild mushrooms have been explored from sub mountainous region of Punjab. Five different basidiomycetesnamely: Pleurotusflorida, Pleurotussajor-caju and Macrolepiotaprocera from order Agaricale, Sparassiscrispa and Trameteselegans from order Polyporales were described morphologically and biochemically. These wild mushrooms were explored during rainy season from two Districtsviz Pathankot with GPS coordination 32°26'43" N 75°64'21" E and Hoshiarpur with GPS coordination 31°51'43.178" N, 75°91'14.83"E. These mushrooms were investigated for their capacity to produce lignocellulosic enzymes. It was observed that endo- β -1,4-glucanase was significantly higher in Trameteselegans(160.14 U/ mg) but xylanase and laccase were found to be lowest in case of Trameteselegans. Total cellulases and cellobiase were found to be higher in Pleurotussajor-caju with values 1.96 U/mg and 1.84 U/mg respectively.

Keywords: Wild Mushroom, Agaricale, Polyporale, Lignocellulosic Enzymes.

1. INTRODUCTION

Mushrooms are indispensible part of the ecosystem. Some mushrooms are cultivated but some are directly collected from the wild and consumed. Among 650-700 edible mushroom species, approximately 130 have been domesticated(Agahar-Murugkarand Subbulakshmi 2005). Wild mushrooms are gaining importance due to their high protein and low fat/energy contents. To combat with world's food shortage problem, mushrooms can occupy a place above vegetables and legumes (Baltacioglu *et al* 2015). Therefore, it is necessary to exploit wild mycoflora for their uses in bioremediation, biodegradation, bio-pesticidal and pharmacological qualities. On the top of everything, it is important to conserve biodiversity of macro-fungi to secure it from the danger of extinction.

India represents diversified agro-climatic zones that cherish a stock of fungal diversity. Though mushroom diversity is very rich in India, 50 % of that is yet to describe (Dwivedi *et al* 2012). So far, about 1,105 to 1,208 species of mushrooms

belonging to 128-130 genera have been documented and among these, 300-315 species belonging to 75-80 genera are considered edible(Kaur*et al* 2015). Punjab harbors in Northeast India represents five different agro-climatic zones namely sub-mountain undulating zone, undulating plain zone, central plain zone, western plain zone and western zone which provides a wide range of opportunity to collect diverse mushroom flora. Its humid climatic conditions, plant distributions and field features are very suitable for the growth of edible fungi.

Furthermore, wild mushrooms provide a scope to degrade cellulosic material as it grows on wide range of substrates because of its capacity to produce lingo cellulosic enzymes *i.e.* cellulases, xyalanases and laccases. These wild mushrooms may serve as alternative path for the use of agrowaste. So,present study was aimed to collect new wild mushrooms and calculate their enzyme producing capacity.

2. MATERIAL AND METHODS

2.1 Survey and collection of Mushrooms

The field survey for collection of various fleshy fungi from sub mountain undulating zone was undertaken during July 2015 to September 2015 and July 2016 to September 2016. Required materials and equipments such as isolation kit, typed performa, digital camera for photography, digging equipment, GPS device were carried along during survey. Photographs of wild mushroom were taken in their natural habitat. The map of collection locality was sketched with GPS coordinate and wild mushrooms were carefully handled, packed in brown paper bags and carried to Mushroom Research Complex, Punjab Agricultural University to record macroscopic features used Standard methods for identification. of collection, preservation, macroscopic and microscopic observations were followed. Characteristics of mushrooms were recorded in structured Performa. Mushroom samples were dried in hot air dryer at 60° C. Some collected edible fleshy fungi were also

cultured on Potato dextrose agar (PDA) medium and maintained for further study.

2.2 Morphological Studies of Collected Edible Mushrooms

All the collected edible mushrooms were observed on the basis of morphological and other phenotypic parameters recorded in fresh samples. The diameter of fruit body of collected mushrooms was measured in centimeters. Length of stalk was also recoded. Presence of different part of fruit bodies like, cap, stalk, gills, volva, annulus, shape and scales were observed. Morphology of stalk, e.g. attachment with cap, shape and its base was also documented. Spore print was prepared by placing the pileus downwards in an inverted jar which was kept on a black paper.

2.3 Raising biomass for production of enzymes

Wild mushroom cultures were grown on mushroom minimal medium (MMM) (*Raper et al* 1972) in 250 ml Erlenmeyer flasks containing 50ml MMM medium for enzyme production. MMM broth was supplemented with wheat straw (@ 0.5 g/flask) as inducers for extracellular enzyme production. All the cultures were grown in triplicate and incubated at 30° C for 10 days. Cultures broth was filtered using whatman paper no. 1 and filtrates were collected for the estimation of enzyme activity.

2.4 Estimation of enzyme activity and total protein

Total cellulases were determined at 50 °C using whatman No.1 filter paper strips (1-6 cm) according to the method of Toyama and Ogawa 1977. Endoglucanase (CMCase) activity

was estimated by the method of Mandels*et al* (1976).The xylanase activity was determined at 50 °C for 10 minutes using beechwoodxylan (1%, w/v, Sigma, USA) as the substrate. The reducing sugar released by the enzymatic reaction was estimated using the DNS method with glucose or xylose as the standard.Assay of laccase was done by the method of Turner (1974). For estimation of total protein, standard method of Lowry *et al* (1951) was followed. Specific enzyme activity is calculated for each enzyme, as the number of enzyme units per mldivided by the concentration of protein in mg/ml. Specific activity values are therefore quoted as units/mg or nmol/min/mg.

3. RESULT AND DISCUSSION

3.1 Morphology of wild mushroom

3.1.1 Pleurotusflorida and Pleurotus Sajor-caju: These mushrooms are saprobic in nature and belongs to the class Basidiomycetes, order Agaricales and familv Tricholomataceae (Table 1). The fruiting bodies were observed on tree trunk (P.florida) and wooden log (P. sajorcaju)grown as a group stacked one over another. The pileus ranges from 6 to 12 cm in diameter. P. florida (Fig.1) and P.sajor-caju (Fig.3) arefan shaped, white in colour. But, at younger stage *P.sajor-caju*is spoon shaped and gives coral like appearance. Stipe is very short and ranges from 1 to 2 cm in length. Gills are running down the stem and spore print is creamy white for P. florida and white for P. sajor-caju. According to Rajarathnam et al (1987) and Ali et al (2012) morphological characters of Pleurotus spp. was described.

| Mushroom | Pleurotusflorida | Macrolepiotaprocera | Pleurotussajor- caju | Sparassiscrispa | Trameteselegans | |
|---|-------------------------|--|--|--|--|--|
| Collection date | 15-08-15 | 3-07-16 | 4-07-16 | 20-07-16 | 2-08-16 | |
| Location, GPS data Gho, Dstt. Pathanko 32o19'45.4"N 75° 40'14.7" E | | Akhwana,Dstt. Pathankot320 16'54" N 75 043'54" E | Gidarpur, Dstt. Pathankot 32011'38.1''N 75° 30'15.2'' E | Dharkalan, Dstt. Pathankot 32o24'18" N 75o47'21" E | Bhangala, Dstt. Hoshiarpur 3201'14"N 75° 36'37" E | |
| Habitat, Vegetation Tree trunk community | | Humus | Dead wooden log | Roots of Malabar tree | Wooden log | |
| Smell, taste Typical mushroom like | | Typical mushroom like | Typical mushroom like | Not distinctive | Not distinctive | |
| Spore print | Spore print Cream | | White | Off white to brown | White | |
| Pileus color | Cream | White | White | Off white to brown | White | |
| Pileus dimension | Pileus dimension 6-8 cm | | 1-3 cm | 10-34 cm | 6-10 cm | |
| Pileus shape, Margin | Fan shaped | Hemispherical to convex type | Spoon shaped | Fan shaped, inrolled margin | semicircular shape | |
| Pileus surface | Smooth | Rough | Smooth | Smooth | tough surface | |
| Scales | No | Present | No | No | No | |
| Stipe attachment | Central | Central | Central | Sessile | Sessile | |
| Stipe size, color Cream,1-2 cm | | Off-white, 10-12 cm | White, 4-5 cm | - | - | |
| Stipe shape, base | Non-bulbous | Bulbous | Non-bulbous | - | - | |
| Ring | Ring No | | No | - | - | |
| Veil | Veil No | | No | - | - | |

Table 1: Morphological study of wild edible mushrooms

| Volva | No | No | No | - | - |
|--------------------------|-------------------|--------|-------------------|--------|---|
| Basal association | No | No | No | - | - |
| Gill attachment | Shortly decurrent | Free | Shortly decurrent | - | - |
| Gill color | Creamish yellow | White | White | - | - |
| Edibility | Edible | Edible | Edible | Edible | - |

3.1.2 *Macrolepiotaprocera*: Mushroom is tall; large in size with pileus dimensions ranges between 6-12 cm (Table 1& Fig. 2). Stipe is longer, 10-12 cm in size, off-white in colour and centrally attached. Base of stipe is bulbous and ring is present. Brown color scales are present on pileus surface. This mushroom is a member of order Agaricale and family Agaricaceae. Gills are crowded, white in colour and attached freely with the cap. Spore print is white in colour. The genus Microlepiota was well described by Geet al (2010).

3.1.3 *Sparassiscrispa*: This mushroom was found on roots of Malabar neem, saprobic in nature (Table 1& Fig. 4). Fruiting bodies are larger in size having diameter 10-34 cm. Pileus is wide composed of tightly packed branches, off-white to yellow in colour. Spore print is white. *Sparassiscrispa* belongs to order Polyporales, family Sparassidaceae. Dictionary of fungi by Kirk *et al* (2010) was referred to describe the collected wild mushrooms.

3.1.4 *Trameteselegans*: This mushroom is tough, white in colour and belongs to order Polyporale family Polyporaceae (Table 1 &Fig. 5). This saprophytic mushroom is semicircular have concentric zone of texture. Cap is 6-10 cm in diameter, found on wooden log andstipe is absent. Pileus has round to angular pores on its surface. Fruit body is very hard and woody, so it is non edible mushroom. Spore print is white in colour. Dictionary of fungi by Kirk *et al* (2010) was referred to describe the collected wild mushrooms.

3.2 Biochemical characterization of wild mushrooms:

It was observed that endo- β -1,4-glucanase was significantly higher in *Trameteselegans*(160.14 U/ mg) but specific activity of xylanaseand laccase was found to be lowest in case of *Trameteselegans*(Table 2).Production of endo- β -1,4-glucanase was lowest in case of *Pleurotus* spp. with specific activity of 14.65 and 7.39 U/mg in *P.florida* and *P. sajor-caju* respectively. Total cellulases and cellobiase were found to be higher in *Pleurotussajor-caju* with values 1.96 U/mg and 1.84 U/mg respectively (Table 2).Goyal and Soni (2011) recorded that activity of endo- β -1,4-glucanase was higher in case of *Pleurotuss*pecies as compare to other enzymes. Laccase activity of *S. crispa* was very low and comparable to the findings of Souet al (2017).

 Table 2: Specific activity (U/mg) of lingo-cellulosic enzymes in wild mushroom culture filtrates.

| S. No. | Mushroo m | Endo-β- 1,4- Glucan ase (U/mg) | Cellobi ase (U/mg) | Total Cellulas es (U/mg) | Xylanas e (U/mg) | Laccass e (U/mg) |
|--------------|-------------------|--|--------------------------|-----------------------------------|-------------------------|-------------------------|
| 1 | P. florida | 14.65 | 0.089 | 0.154 | 0.724 | 2.14 |
| 2 | M. procera | 42.91 | 0.91 | 0.87 | 12.53 | 1.38 |
| 3 | P. sajor- caju | 7.39 | 1.84 | 1.96 | 3.03 | 0.87 |
| 4 | S. crispa | 83.84 | 0.320 | 0.228 | 2.08 | 0.273 |
| 5 | T. elegans | 160.14 | 0.058 | 0.144 | 0.07 | 0.08 |
| LSD = 0.05 % | | 1.74 | 0.257 | 0.113 | 0.148 | 0.304 |

4. ACKNOWLEDGEMENTS

The authors are thankful to the Punjab Agricultural University, Ludhiana for providing funds and facility for field and laboratory work.

REFERENCES

- [1] Aghar-Murugkar D and Subbulakshmi G (2005) Nutritional value of edible wild mushrooms collected from the Khasi hills of Meghalya. *Food Chem***89**: 599-03.
- [2] Ali B D, KhanA U and Musa H (2012) Studies on the morphology and taxonomyof some species of Tricholomataceae in Zaria, Kaduna State, Nigeria. *J BiolSci Bio-Cons* 4: 13-25.
- [3] Baltacioglu H, Bayndrl A, Severcan M and Severcan F (2015) Effect of thermal treatment on secondary structure and conformational change of mushroom polyphenol oxidase (PPO) as food quality related enzyme: A FTIR study. *Food Chem*187: 263-269
- [4] Dwivedi S, Tiwari M K, Chauhan U.K and Pandey A.K. (2012) Biodiversity of mushrooms of Amarkantak Biosphere Reserve forest of Central India. International Journal of Innovative Research in Science, Engineering and Technology. International Journal Of Pharmacy & Life Sciences. 3(1)
- [5] Ge Z W and Yang Zhu L and Else C (2010) The genus Macrolepiota (Agaricaceae, Basidiomycota) in China Vellinga. *Fungal Diversity* 45:81–98.
- [6] GoyalMeenakshi and Soni Giridhar (2011) Production and characterization of cellulolytic enzymes by *Pleurotusflorida*. *African J Microbiol Res*5(10): 1131-1136.
- [7] Kaur A, Atri N S and Kaur M (2015). Taxonomic study on the coprophilous mushrooms from Punjab, India: new records of family Agaricaceae. *Current Res Environ & Applied Mycology*5: 27-45.

- [8] Kirk P M, Cannon P F, Minter D W, Stalpers J A (2008) Dictionary of the Fungi.10th edn. CAB International, Wallingford, UK.
- [9] Lowry O H, Rosebrough N J, Farr A L and Randall R J (1951) Protein measurements with Folin Phenol regeant. J BiolChem193:265-75.
- [10] Mandels M, Andreotti R and Roche C (1976). Measurements of saccharifying cellulose. *BiotechnolBioenggSymp*. 6:21-23
- [11] Poonambalam A S, Deepthi R S, Ghosh A R (2011) Qualitative display and measurement of enzyme activity of isolated cellulytic Research article. *BiotechnolBioinfBioeng*1: 33-37
- [12] RajarathnamS, BanoZakia and Miles Philip G (1987) *Pleurotus* mushrooms. Part I A. morphology, life cycle, taxonomy, breeding, and cultivation. *Critical Reviews Food Sci&Nutri* 26 (2):157-223.
- [13] Raper C A, Miller R E and Raper J R (1972).Genetic analysis of the life cycle of *Agaricusbisporus*. Mycologia64: 178-84.
- [14] Sou Hong-Duck, RyooRhim, Kab Kang-Hyeon and Park Hyun (2017) The mycelial growth and ligninolytic enzyme activity of cauliflower mushroom (*Sparassislatifolia*). Forest Sci& Tech13(4): 158-163.
- [15] Toyama M and Ogawa K (1977) Cellulose production of *Trichodermaviridae* in solid and submerged culture methods. In Ghosh T K (ed).,ProcSymp on Bioconversion of cellulosic substrates into Energy, Chemicals and Microbial Protein. IIT Delhi, India 7: 305-12
- [16] Turner E M (1974) Phenoloxidase activity in relation to substrate and devolpmental stage in mushroom *Agaricusbisporus. Trans Br MycolSoc***63**: 541-47.



Figure 1: Pleurotusflorida



Figure 2: Macrolepiotaprocera



Figure 3: Pleurotussajor-caju



Figure 4: Sparassiscrispa



Figure 5: Tarmeteselegans